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Review

Peripheral tachykinin receptors as targets for new drugs

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Abstract

Tachykinins are widely distributed in the peripheral nervous system of the respiratory, urinary and gastrointestinal tract, stored in enteric neurons and in peripheral nerve endings of capsaicin-sensitive primary afferent neurons from which are released by stimuli having both pathological and physiological relevance. The most studied effects produced by tachykinins in these systems are smooth muscle contraction, plasma protein extravasation, mucus secretion and recruitment/activation of immune cells. The use of tachykinin receptor-selective antagonists and knockout animals has enabled to identify the involvement of tachykinin NK₁, NK₂ and NK₃ receptors as mediators of peripheral effects of tachykinins in different systems/species. The bulk of data obtained in experimental animal models suggests that tachykinins could contribute to the genesis of symptoms accompanying various human diseases including asthma/bronchial hyperreactivity, cystitis of various aetiology, inflammatory bowel diseases and irritable bowel syndrome. Tachykinin receptor antagonists are expected to afford therapeutically relevant effects. © 2001 Elsevier Science B.V. All rights reserved.

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1. Introduction

1.1. Mammalian tachykinins

Tachykinins are a family of peptides which share the common C-terminal sequence Phe-Xaa-Gly-Leu-Met-NH₂. This sequence is crucial for their interaction with specific receptors and for producing the most of their biological effects. However, other domains of the tachykinin peptide sequence have been recognized to be determinant for producing certain effects (e.g. mast cell degranulation encoded by the N-terminal sequence of substance P) (see Maggi et al., 1993, for review). The first peptide of this family, substance P, was discovered by Von Euler and Gaddum as early as in 1931; more than 50 years elapsed before two other mammalian tachykinins, neurokinin A and neurokinin B, were disclosed (Maggio, 1988, for review). To date, substance P, neurokinin A and neurokinin B are three mammalian tachykinins that have reached an established status of neurotransmitters. Two genes have been identified in mammals encoding peptides of the tachykinin family: the preprotachykinin I gene (PPT-I or PPT-A) which encodes both substance P and

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neurokinin A, and the preprotachykinin II gene (PPT-II or PPY-B) which encodes neurokinin B. The primary RNA transcript of the PPT-I gene is spliced to yield four different forms of messenger ribonucleic acids (mRNA) termed α , β , γ and δ forms (Nawa et al., 1983, 1984; Krause et al., 1987; Nakanishi, 1987). α and δ PPT-I mRNAs code for the synthesis of substance P whereas β and γ PPT-I mRNAs code for the synthesis of both substance P and neurokinin A. In addition, β and γ PPT-I mRNAs encode the synthesis of two N-terminally extended forms of neurokinin A (known as neuropeptide K and neuropeptide γ, respectively): both peptides are capable to produce full biological responses (e.g. Takeda and Krause, 1989a,b) but their role as neurotransmitters awaits to be proven. On the other hand, the PPT-II gene encodes neurokinin B only (Maggio, 1988, for review). The PPT-I gene has been detected in both central and peripheral nervous system, in enteric neurons of the gut and in various cells of the immune system. In contrast, the PPT-II gene is expressed almost exclusively in the central nervous system (Maggi et al., 1993; Maggi, 1995 for review). Recently, the molecular cloning of a further preprotachykinin gene (termed PPT-C) has been described from cDNA of murine hematopoietic cells (Zhang et al., 2000). PPT-C mRNA was detected in pro- and pre-B lymphocyte cells from bone marrow, whereas it was undetectable in nonhematopoietic cells. The novel tachykinin encoded by PPT-C gene (termed

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hemokinin I) may act as an autocrine factor for the growth of hematopoietic cells. Zhang et al. (2000) have also suggested that hemokinin I may stimulate a specific receptor, distinct from any other known tachykinin receptor. However, further experimentation is needed to clearly define the role of hemokinin I in mammals. Owing to the lack of PPT-II gene expression in peripheral tissues, substance P and neurokinin A are the only tachykinins detected in appreciable amounts at this level. In normal, noninflamed tissues, tachykinins originate from neuronal cells, being present in (i) peripheral endings of capsaicinsensitive primary afferent neurons (Maggi, 1995, for review) and (ii) enteric neurons of both submucousal and myenteric plexuses innervating all layers of the gut (Holzer and Holzer-Petsche, 1997a,b, for review). In addition, certain immune cell types synthesize and possibly release tachykinins during inflammation, thus representing a nonneuronal source of releasable tachykinins in inflamed tissues (De Giorgio et al., 1998; Lai et al., 1998; Maggi, 1997 for review).

1.2. Mammalian tachykinin receptors

The existence of multiple tachykinin receptors was first suggested by the observation that several nonmammalian tachykinins (i.e. eledoisin and kassinin) possess much higher potency than substance P in certain mammalian isolated tissues (Erspamer, 1981). Following the discovery of neurokinin A and neurokinin B in the early 1980s, several groups have presented evidence, based on functional or radioligand binding data, for the existence of three distinct receptors (termed NK₁, NK₂ and NK₃) mediating the biological actions encoded by the common C-terminal sequence of tachykinins. Substance P, neurokinin A and neurokinin B were initially considered to be the "preferred" ligands for the tachykinin NK₁, NK₂ and NK₃ receptors, respectively. (see Regoli et al., 1989; Guard and Watson, 1991; Mussap et al., 1993; Maggi et al., 1993; Maggi, 1995 for reviews). However, this concept has been challenged by radioligand binding and functional experiments that have shown both neurokinin A and neurokinin B to be as potent as substance P at stimulating the tachykinin NK₁ receptor (Hastrup and Schwartz, 1996; Maggi and Schwartz, 1997). It is also worth-noting that all three mammalian tachykinins (substance P, neurokinin A and neurokinin B) are capable to act as full agonists on each one of the three receptors, albeit at different concentrations. In 1996, the cloning of a further tachykinin receptor showing a high degree of structural homology and a similar binding profile to the previously identified NK₃ receptor protein has been reported (Donaldson et al., 1996; Krause et al., 1997). However, a recent attempt to isolate this receptor (referred to as "NK₄" or "NK_{3B}" in literature) or identify the corresponding gene by the use of various molecular biological techniques in human and other species was unsuccessful (Sarau et al., 2000). In

view of these latter results, any update of the current tachykinin receptor classification aiming at including the "NK₄" receptor seems to be premature. Likewise, the possible existence of a hemokinin I-operated tachykinin receptor, as proposed by Zhang et al. (2000), needs to be confirmed in further investigations. The tachykinin NK₁, NK₂ and NK₃ receptors belong to the superfamily of rhodopsin-like, G-protein-coupled receptors with seven transmembrane spanning segments. Stimulation of phosphoinositol breakdown is a common effector system coupled to each one of the three tachykinin receptors, although other intracellular pathways are triggered upon occupation of tachykinin receptors as well, e.g. leading to cAMP formation and increased arachidonic acid metabolism (Henderson et al., 1990; Eistetter et al., 1991, 1993; Arkinstall et al., 1994; Catalioto et al., 1998b). Marked speciesrelated differences in pharmacology exist for all three tachykinin receptors, revealed by the use of selective tachykinin receptor antagonists (see Patacchini and Maggi, 1995; Maggi, 1995 for review). Stimulation of peripheral tachykinin receptors leads to specific biological effects including: smooth muscle contraction, neuronal stimulation, endothelium-dependent vasodilation, plasma protein extravasation, chemotaxis and activation of immune cells, stimulation of secretion (Maggi, 1995, for review). The introduction of the first potent and selective tachykinin receptor antagonists at the beginning of the 1990s has allowed to assess the (patho)-physiological role of tachykinins in the central and peripheral nervous system. Either preclinical and clinical trials have shown that centrally acting tachykinin NK₁ receptor antagonists are endowed with anti-depressant, anxyolitic and anti-emetic therapeutic potential (Rupniak and Kramer, 1999; for review). On the other hand, clinical evidence supporting the use of tachykinin antagonists acting at peripheral level for treatment of human diseases is not available yet. Nevertheless, there is a general expectation that tachykinin receptor antagonists may prove useful for treatment of a variety of human diseases at visceral level. This review is focused on the pathophysiological role played by tachykinins in the respiratory, genitourinary and gastrointestinal systems, and on the preclinical evidence indicating a possible therapeutic use of antagonist compounds acting at peripheral tachykinin receptors.

2. Airways

Since the first demonstration that tachykinins are the sensory neuropeptides released by capsaicin from primary afferent nerve endings innervating the guinea-pig airways to produce smooth muscle contraction (Szolcsanyi and Barthò, 1982; Lundberg and Saria, 1982) and that tachykinin-containing nerve fibres are present in normal, but not in capsaicin-pretreated animals (e.g. Lundberg and Saria, 1982; Lundberg et al., 1983), a large number of studies

have been undertaken to assess the hypothesis that tachykinins may be involved in airways pathophysiology. Taken as a whole, these investigations have suggested that tachykinins, released from capsaicin-sensitive sensory nerves innervating upper and lower airways of several species including man (Lundberg et al., 1984; Martling et al., 1988), may participate in the genesis of asthma/ bronchial hyperreactivity and rhinitis. Tachykinins have been shown to produce a variety of responses at respiratory level, including tracheal/bronchial smooth muscle contraction in man and other species (e.g. Maggi et al., 1991a,b; Naline et al., 1989), stimulation of secretions in both upper (e.g. Geppetti et al., 1988; Petersson et al., 1989) and lower (e.g. Coles et al., 1984; Meini et al., 1993) respiratory tract, development of bronchial hyperreactivity following exposure to inflammatory agents (antigens and others) (e.g. Saria et al., 1983; Boichot et al., 1995; Schuiling et al., 1999), plasma protein extravasation (e.g. Saria et al., 1983), recruitment of inflammatory cells and stimulation of resident immune cells (e.g. alveolar macrophages; Brunelleschi et al., 1990; Maggi, 1997, for review), sneezing (e.g. Stjärne et al., 1989) and nasal obstruction (Devillier et al., 1988). It is worth-mentioning that most stimuli capable of releasing tachykinins from sensory nerves in the airways are of pathological rather than physiological relevance. Evidence for tachykinin release from peripheral primary afferent terminals has been provided for the following: (a) administration of antigens to sensitized animals, (b) inhalation of irritants (like capsaicin, cigarette smoke, acid media, cold air, occupational sensitizers as toluene diisocyanate and others), (c) administration of pro-inflammatory autacoids like bradykinin and histamine (Maggi, 1995; Advenier et al., 1999; Kraneveld et al., 2000; Lecci et al., 2000).

2.1. Tachykinin receptors mediating pathophysiological effects of tachykinins in the airways

The introduction of tachykinin receptor selective antagonists since the middle 1980s has allowed to investigate more properly the relative contribution of each tachykinin receptor type to the biological effects produced by tachykinins at respiratory level. Bronchoconstriction of human airways caused by tachykinins is an effect mediated by tachykinin NK₂ receptors only (Naline et al., 1989; Dion et al., 1990; Ellis et al., 1993), while tachykinin NK₁ receptors are partially contributing to this effect in other species (e.g., Maggi et al., 1991b). Plasma protein extravasation and mucus secretion are two other effects mainly (if not exclusively) mediated by tachykinin NK₁ receptors in all species examined (Maggi, 1995; Lecci et al., 2000, for reviews). Recruitment and activation of inflammatory leukocytes induced by tachykinins are further effects mainly mediated by tachykinin NK₁ receptors (Advenier et al., 1997; Maggi, 1997), although stimulation of tachykinin NK₂ receptors produces infiltration of neutrophils and lymphocytes in airways of ovoalbumin-challenged guineapigs (Schuiling et al., 1999).

2.2. Clinical trials in human airways

Only few clinical studies are currently available, describing tachykinin receptor antagonists challenged to counteract asthmatic reactions provoked by different stimuli. In the first of these reports, Ichinose et al. (1992) claimed the effectiveness of the dual (NK₁/NK₂) tachykinin receptor antagonist (FK 224), in preventing both cough and bronchoconstriction elicited by inhaled bradykinin in asthmatic patients. However, further investigations with FK 224 failed to prove this compound capable of preventing neurokinin A-induced bronchoconstriction in asthmatics (Joos et al., 1996) or to ameliorate symptoms/lung function in patients suffering of mild to moderate asthma (Lunde et al., 1994). The use of a tachykinin receptor antagonist selective for the NK₁ receptor, the nonpeptide compound (CP 99994), did not afford better results, since CP 99994 (intravenously administered to mild asthmatic patients) failed to prevent bronchoconstriction and cough induced by hypertonic saline (Fahy et al., 1995). Likewise, the tachykinin NK₁ receptor-selective antagonist FK 888 failed to afford beneficial effects on exercise-induced bronchoconstriction in asthmatics, but significantly shortened the recovery times (Ichinose et al., 1996), suggesting a possible role of tachykinin NK₁ receptors in the late bronchoconstrictor response to exercise. On the other hand, the tachykinin NK2 receptor-selective antagonist (SR 48968, saredutant; Emonds-Alt et al., 1992) is the first compound proven active (by oral route) in reducing bronchoconstriction provoked by inhaled neurokinin A in mild asthmatic patients (Van Schoor et al., 1998). The capability of tachykinin NK2 receptor-selective antagonists to inhibit neurokinin A-induced bronchoconstriction in asthmatics has been further demonstrated by the use of nepadutant (Catalioto et al., 1998a) in a double-blind, placebo controlled cross-over trial (Joos et al., 2000). Nevertheless, a recent study (Kraan et al., 2001) has reported that saredutant (100 mg once a day, per os) was unable to reduce airways hyperresponsiveness caused by adenosine 5'-monophosphate in allergic asthmatic patients. Further trials are awaited to prove efficacy of tachykinin receptor blockers in asthmatic responses caused by allergens producing bronchoconstriction, bronchial hyperreactivity and airways inflammation.

3. Urinary tract

All the organs of upper and lower urinary tract of various species, including man, are densely innervated by peripheral projections of capsaicin-sensitive primary afferent neurons, which represent the main if not the sole source of tachykinins at this level. (Hua et al., 1987;

Edyvane et al., 1992; Edyvane and Marshall, 1990). Tachykinins can be released from capsaicin-sensitive primary afferent neurons in the urinary tract by a number of different stimuli, having pathological rather than physiological relevance including: acid media (Geppetti et al., 1990), various irritants like xylene (Maggi et al., 1988a) and cyclophosphamide (Maggi et al., 1992a), products of bacterial metabolism (Giuliani et al., 1991), certain constituents of urine like K⁺ (Maggi et al., 1989b) and bradykinin (Maggi et al., 1989a).

3.1. Tachykinin receptors mediating pathophysiological effects of tachykinins in the urinary tract

Smooth muscle contraction is one of the most studied effects produced by tachykinins throughout the urinary tract. The urinary bladder detrusor muscle and urethra of various species (man included) respond with a contraction to exogenously applied tachykinins while, in selected species, nerve-mediated urinary bladder contractions produced by endogenous tachykinins can be observed upon application of capsaicin or electrical nerve stimulation (Maggi, 1995, for review). Tachykinin NK₁ and/or NK₂ receptors, working alone or in combination, are the mediators of tachykinin-induced contractions in animal species. In man the tachykinin NK, receptor exclusively mediates tachykinin-induced urinary bladder contraction (Maggi et al., 1988b), and it is also present in the urethra (Parlani et al., 1990; Palea et al., 1996) and ureter (Patacchini et al., 1998). In the spontaneously active guinea-pig renal pelvis tachykinins, exogenously applied or released by capsaicin or electrical nerve stimulation, produce both inotropic and chronotropic effects (Maggi et al., 1992b). Plasma protein extravasation is an acute inflammatory response that is produced by exogenously applied tachykinins in organs of the urinary tract, or by agents that stimulate the release of tachykinins from sensory nerves, like capsaicin (Eglezos et al., 1991), xylene (Giuliani et al., 1993b), cyclophosphamide (Ahluwalia et al., 1994) or antigens in sensitized animals (e.g. Ahluwalia et al., 1998) (Lecci and Maggi, 1995, for review). As in the airways, tachykinin-mediated plasma protein extravasation in the urinary tract requires activation of tachykinin receptors of the NK₁ type. The involvement of tachykinin NK₁ receptors in mediating this effect was proven since the introduction of the first receptor-selective agonists and antagonists (e.g. Eglezos et al., 1991). Recently, the involvement of the NK₁ receptor has been confirmed by the failure of cyclophosphamide to produce plasma protein extravasation in the urinary bladder of NK₁ receptor knockout mice (Laird et al., 2000). Owing to the ability of exogenously administered tachykinins to stimulate ureteral peristalsis and activate the micturition reflex (Maggi, 1995; Santicioli and Maggi, 1998), it could be hypothesized that endogenous tachykinins, released into the bladder wall, take part into the voiding mechanism of the urinary bladder. However, Lecci et al. (1993, 1998b) provided evidence against a peripheral role of tachykinins in initiating volume-evoked micturition reflex in rats, by showing that peripherally acting tachykinin NK1 and/or NK2 receptor selective antagonists are unable to modify the urodynamic parameters of distension-induced micturition under physiological conditions. By contrast, endogenous tachykinins acting via NK₂ receptors contribute to detrusor hyperreflexia generated by noxious stimuli, as shown by the ability of tachykinin NK₂ (but not NK₁) receptor-selective antagonists to correct altered urodynamic parameters (micturition frequency and bladder tone) following intravesical administration of irritant agents in rats (like capsaicin, prostaglandin E2, xylene, E. coli lipopolysaccharide,) (Pietra et al., 1992; Ishizuka et al., 1994, 1995; Lecci et al., 1997, 1998c). To this regard, it may be speculated that tachykinins mediate chemically induced hyperreflexia by directly stimulating afferent nerve fibers in the mucosal layers of the bladder, on which are expressed specific (NK₂) receptors. Alternatively, tachykinins might act indirectly by stimulating generation of other local transmitters like prostanoids (see Lecci et al., 2000 for discussion). The bulk of data collected in in vitro and in vivo studies on animal models suggests that tachykinins could be involved in producing symptoms accompanying cystitis of different aetiology. Nonetheless, the therapeutical potential of tachykinin receptor antagonists in human diseases of the urinary tract has not yet been investigated.

4. Gut

The main source of tachykinins in the gastrointestinal tract is represented by intrinsic enteric neurons, including cholinergic motoneurons localized in the myenteric plexus and projecting to circular and longitudinal smooth muscle layers. The rest of tachykinin content is contributed by extrinsic fibres of capsaicin-sensitive primary afferent neurons, and by immune cells (Costa et al., 1987; Maggi, 1995, 1997; Holzer and Holzer-Petsche, 1997a). Tachykinins stored in primary afferent nerves are released by the same stimuli, mainly of irritative nature, which induce their release in the airways and urinary tract. Tachykinins stored in enteric neurons can be released by mechanical (i.e. intestinal wall distension) or chemical (synaptic input) stimuli having a physiological relevance (Maggi, 1995; Holzer and Holzer-Petsche, 1997a, for review).

4.1. Tachykinin receptors mediating pathophysiological effects of tachykinins in the gut

As seen in the organs belonging to respiratory or urinary tract, tachykinins are potent spasmogens in almost all intestinal regions so far investigated. A large part of their contractile effects originates from direct stimulation of tachykinin NK_1 and/or NK_2 receptors present on smooth

muscle cells/cells of Cajal of the circular and longitudinal muscle layers (Holzer and Holzer-Petsche, 1997a). In addition, indirect contractile responses can be evoked by tachykinins through stimulation of tachykinin receptors (mainly of the NK₃ type) present on intestinal neurons, from which either acetylcholine or tachykinins themselves are released (Patacchini et al., 2000). It is worth-mentioning that the release of inhibitory transmitters (e.g. nitric oxide) has also been reported upon stimulation of tachykinin NK₁, NK₂ and NK₃ receptors (Maggi et al., 1997). A great number of studies have reported that both direct/ indirect excitatory motor responses elicited in mammalian intestine by application of electrical or mechanical (i.e. intestinal wall distension) stimuli, are mediated by endogenous tachykinins which add to the major effective excitatory transmitter, acetylcholine (Maggi, 1995; Holzer and Holzer-Petsche, 1997b; Maggi et al., 1997 for review). In this respect, it is noteworthy that the contribution of endogenous tachykinins to intestinal peristalsis is hardly appreciable under normal conditions, unless muscarinic receptors are blocked. Giuliani et al. (1993a, 1996) have shown that in atropine-pretreated animals, distensionevoked intestinal reflex contractions can be elicited and that tachykinin NK₁ and NK₂ receptor-selective antagonists inhibit these atropine-resistant responses, proving the involvement of endogenous tachykinins. Similar results were obtained by Lecci et al. (1998a) studying colonic propulsive activity in anaesthetized atropine-pretreated guinea-pigs, and by Holzer et al. (1998) studying in vitro peristalsis of isolated segments of guinea-pig small intestine. However, the former studies have also proven that tachykinins do not play a major role in intestinal motor activity under normal conditions, due to the modest/null ability of tachykinin receptor antagonists to modify normal peristalsis. This concept would be supported by recent studies performed in unanaesthetized dogs (Giuliani et al., 2001) and in healthy volunteers (Lördal et al., 1999) in which the tachykinin NK₂ receptor-selective antagonist nepadutant (Catalioto et al., 1998a) did not affect gastrointestinal normal motor activity per se. However, Tonini et al. (2001) showed that simultaneous administration of three tachykinin receptor antagonists, each one selective for the NK₁, NK₂ or NK₃ receptor type, produced a 50% inhibition of velocity of propulsion of a balloon inserted in distal colon isolated segments from guinea-pig. More convincing evidence exists on the role of endogenous tachykinins in producing exaggerated intestinal motility associated with various inflammatory and infectious intestinal diseases (Holzer, 1998 for review). In this context, tachykinin NK₂ (and partially NK₁) receptor antagonists have been reported to prevent increased fecal excretion and colonic giant contraction in castor-oil induced diarrhea in rats (Croci et al., 1997), without producing constipation. Besides excitatory motor responses, inhibitory motor effects have been reported following stimulation of tachykinin NK₁, NK₂ or NK₃ receptors (Maggi et al.,

1997; Lecci et al., 2000). Either stimulation of tachykinin (NK₂) receptors present on sympathetic ganglia and subsequent activation of inhibitory sympathetic nerves (Giuliani et al., 1988), or activation of tachykinin (NK₁, NK₂ or NK₃) receptors present on enteric inhibitory neurons which in turn release nitric oxide, have been proposed as possible mechanisms of tachykinin-induced inhibitory motor effects at intestinal level (Holzer and Holzer-Petsche 1997b; Lecci et al., 1999; Zagorodnyuk and Maggi, 1995). To this regard, it may be speculated that tachykinin receptor antagonists may result useful in abnormal (inhibited) intestinal motility, as the post-surgical ileus. As a matter of fact, preliminary evidence has been obtained indicating that the tachykinin NK, receptor antagonist nepadutant (Catalioto et al., 1998a,b) reduces the inhibition of jejunal motility induced by abdominal surgery in rats (Bueno et al., unpublished observations). Besides affecting smooth muscle motility of the gut, tachykinins have been reported to enhance electrolyte and/or fluid secretion from both small and large intestine. At this level tachykinins would act as endogenous modulators by acting on secretomotor neurons which in turn release acetylcholine and other noncholinergic transmitters which in turn cause ion and fluid secretion (Maggi, 1995; Holzer and Holzer-Petsche, 1997b for review). For example, Cox et al. (1993) showed that in the rat descending colon mucosa under voltage-clamp conditions, tachykinins evoked increases in short-circuit current (I_{sc}) responses by activating a combination of NK₁, NK₂ and NK₃ receptor types. In a recent study, Patacchini et al. (2001) showed that in the rat distal colon muscularis mucosae, tachykinin NK2 receptor-mediated contractile responses are produced by activating an indomethacininsensitive effector mechanism, whereas tachykinin NK, receptor-mediated ion transport in the colon mucosa is a largely indomethacin-sensitive phenomenon. These results along with the observation that nepadutant blocks with different kinetics (competitive vs. noncompetitive, respectively) the two responses, provide indirect evidence that different intracellular effector mechanisms are triggered by the tachykinin NK₂ receptor in the intestine and lead to different biological responses.

Several animal models of ileocolitis are presently available, in which inflammatory reactions are obtained by administration of chemical irritants (like trinitrobenzensulfonic acid, TNBS, dextran sulphate, formalin), γ -irradiations, bacterial infection (e.g. *Trichinella spiralis, Salmonella, Nyppostrongylus brasiliensis*) or bacterial toxins (e.g. *Clostridium difficile* toxin A and *T. spiralis* toxin A). Endogenous tachykinins have been reported to mediate hypersecretory and inflammatory reactions of the gut induced by these agents, while administration of tachykinin NK₁ receptor antagonists has been shown to exert protective effects in various models of intestinal injury/inflammation (Holzer, 1998; Evangelista, 2001; Lecci et al., 2000 for reviews). The role of the tachykinin NK₁ receptor in mediating inflammatory reactions of the gut has recently

been supported by the use of tachykinin NK₁ receptorknockout mice, which showed reduced intestinal damage produced by C. difficile toxin A or TNBS compared to control animals (Castagliuolo et al., 1998). In addition to mediating abnormal motility and inflammatory reactions in various experimental pathological conditions of the gut, tachykinins have also been shown to take part in visceral nociception (Holzer, 1998, for review). In particular, tachykinin NK₂ receptor antagonists have been found effective in reducing reflex responses (e.g. abdominal contractions) caused by painful stimuli as rectal wall distension (Julia et al., 1994) or intraperitoneal administration of acetic acid (Julia and Bueno, 1997) in rats. Moreover, tachykinin NK₂ receptor antagonists have been proven to be effective in animal models of visceral hyperalgesia induced by inflammation (e.g. McLean et al., 1997) or stress (Toulouse et al., 2000). The results obtained by Toulouse et al. (2000) showing the ability of nepadutant to inhibit rectal hypersensitive responses in rats pretreated with TNBS or previously subjected to restraint, suggest that the tachykinin NK₂ receptor is a main target mediating visceral allodynia/hyperalgesia. The role of tachykinin NK₂ receptors is further supported by the observation that nepadutant prevents the increased expression of either c-fos and c-jun protooncogene markers in spinal cord and dorsal root ganglia neurons from rats pretreated with TNBS (Kiss et al., 1999) and by Laird et al. (in press), who have shown nepadutant capable of preventing the hypersensitivity of single spinal cord neurons responding to colorectal distension or pelvic nerve stimulation in rats pretreated with intracolonic acetic acid. It is worth-mentioning that nepadutant had no significant effect on responses evoked in animals with noninflamed colon (Laird et al., in press).

The precise site at which tachykinin NK₂ receptors mediate visceral pain is yet to be identified, although the peripheral (vs. central) level seems more likely for several reasons (Laird et al., in press; Holzer, 1998). Nevertheless, the effectiveness of tachykinin NK₂ receptor antagonists in animal models of visceral hyperalgesia leads to speculate that these agents could be useful for the treatment of functional gastrointestinal disorders characterized by visceral pain as irritable bowel syndrome.

5. Conclusions

A huge amount of preclinical data indicates that both tachykinin NK₁ and NK₂ receptors are consistently expressed in the peripheral nervous system of several animal species, including humans. Tachykinin NK₃ receptors are also expressed in the peripheral nervous system of certain species, but their role in human peripheral nervous system is much less documented. The results obtained with selective receptor antagonists in animal models of various diseases indicate that tachykinin receptor antagonists might

have a potential therapeutic effect in asthma/bronchial hyperreactivity, cystitis, inflammatory bowel disease and irritable bowel syndrome. To test this hypothesis several compounds are presently under clinical investigation.

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References

- Advenier, C., Lagente, V., Boichot, E., 1997. The role of tachykinin receptor antagonists in the prevention of bronchial hyperresponsiveness airway inflammation and cough. Eur. Respir. J. 10, 1892–1906.
- Advenier, C., Joos, G., Molimard, M., Lagente, V., Pauwels, R., 1999.Role of tachykinins as contractile agonists of human airways in asthma. Clin. Exp. Allergy 29, 579–584.
- Ahluwalia, A., Maggi, C.A., Santicioli, P., Lecci, A., Giuliani, S., 1994. Characterization of the capsaicin-sensitive component of cyclophosphamide induced inflammation in the rat urinary bladder. Br. J. Pharmacol. 111, 1017–1022.
- Ahluwalia, A., Giuliani, S., Scotland, R., Maggi, C.A., 1998. Ovoalbumin-induced neurogenic inflammation in the bladder of sensitized rats. Br. J. Pharmacol. 124, 190–196.
- Arkinstall, S., Emergy, I., Church, D., Chollet, A., Kawashima, E., 1994.Calcium influx and protein kinase C activation mediates arachidonic acid mobilization by the human neurokinin 2 receptors expressed in Chinese hamster ovary cells. FEBS Lett. 338, 75–80.
- Boichot, E., Germain, N., Lagente, V., Advenier, C., 1995. Prevention by the tachykinin NK₂ receptor antagonist, SR 48968, of antigen-induced airway hyperresponsiveness in sensitized guinea-pigs. Br. J. Pharmacol. 114, 259–261.
- Brunelleschi, S., Vanni, L., Ledda, F., Giotti, A., Maggi, C.A., Fantozzi, R., 1990. Tachykinins activate guinea-pig alveolar macrophages: involvement of NK₂ and NK₁ receptors. Br. J. Pharmacol. 100, 417–420.
- Castagliuolo, I., Pasha, C., Wang, C., Keates, A.C., Nikulasson, S., Lu, B., Gerard, N.P., Pothoulakis, C., 1998. Diminished intestinal inflammatory and secretory responses in substance P (SP) receptor (NK-1R) deficient mice. Gastroenterology 114, A1133.
- Catalioto, R.-M., Criscuoli, M., Cucchi, P., Giachetti, A., Giannotti, D., Giuliani, S., Lecci, A., Lippi, A., Patacchini, R., Quartara, L., Renzetti, A.R., Tramontana, M., Arcamone, F., Maggi, C.A., 1998a. MEN 11420 (Nepadutant), a novel glycosylated bicyclic peptide tachykinin NK₂ receptor antagonist. Br. J. Pharmacol. 123, 1–91.
- Catalioto, R.-M., Cucchi, P., Renzetti, A.R., Criscuoli, M., Maggi, C.A., 1998b. Independent coupling of the human tachykinin NK₂ receptor to phospholipases C and A₂ in transfected Chinese hamster ovary cells. Naunyn-Schmiedeberg's Arch. Pharmacol. 358, 395–403.
- Coles, S.J., Neill, K.H., Reid, L.M., 1984. Potent stimulation of glycoprotein secretion in canine trachea by SP. J. Appl. Physiol. 57, 1323–1327.
- Costa, M., Furness, J.B., Llewellyn-Smith, I.J., 1987. Histochemistry of the enteric nervous system. In: Johnson, L.R. (Ed.), Physiology of the Gastrointestinal Tract. Raven Press, New York, NY, pp. 1–40.
- Cox, H.M., Tough, I.R., Grayson, K., Yarrow, S., 1993. Pharmacological characterization of neurokinin receptors mediating anion secretion in rat descending colon mucosa. Naunyn-Schmiedeberg's Arch. Pharmacol. 348, 172–177.
- Croci, T., Landi, M., Emonds-Alt, X., Le Fur, G., Maffrand, J.P.,

- Manara, L., 1997. Role of tachykinins in castor oil diarrhoea in rats. Br. J. Pharmacol. 121, 375–380.
- De Giorgio, R., Tazzari, P.L., Barbara, G., Stanghellini, V., Corinaldesi, R., 1998. Detection of substance P immunoreactivity in human peripheral lymphocytes. J. Neuroimmunol. 82, 175–181.
- Devillier, P., Dessangies, J.F., Rakotosihanaka, F., Ghaem, H.A., Boushey, H.A., Lockhart, A., Marsac, J., 1988. Nasal response to SP and methacoline in subjects with and without allergic rhinitis. Eur. Respir. J. 1, 356–361.
- Dion, S., Rouissi, N., Nantel, F., Drapeau, G., Regoli, D., Naline, E., Advenier, C., 1990. Receptors for neurokinins in human bronchus and urinary bladder are of the NK-2 type. Eur. J. Pharmacol. 178, 215–219.
- Donaldson, L.F., Haskell, C.A., Hanley, M.R., 1996. Functional characterization by heterologus expression of a novel cloned tachykinin peptide receptor. Biochem. J. 320, 1–5.
- Edyvane, K.A., Marshall, V.R., 1990. Neuropeptides in the human urinary tract. Neurourol. Urodyn. 9, 346–347.
- Edyvane, K.A., Trussell, D.C., Jonavicius, J., Henwood, A., Marshall, V.R., 1992. Presence and regional variation in peptide-containing nerves in the human ureter. J. Auton. Nerv. Syst. 39, 127–137.
- Eglezos, A., Giuliani, S., Viti, G., Maggi, C.A., 1991. Direct evidence that capsaicin-induced plasma protein extravasation is mediated through tachykinin NK₁ receptors. Eur. J. Pharmacol. 209, 277–279.
- Eistetter, H.R., Church, D.J., Mills, A., Godfry, P.P., Capponi, A.M., Brewster, R., Schulz, M.F., Kawashima, E., Arkinstall, S.J., 1991. Recombinant bovine neurokinin-2 receptor stably expressed in chinese hamster ovary cells couples to multiple signal transduction pathways. Cell Regul. 2, 779–787.
- Eistetter, H.R., Mills, A., Arkinstall, S.J., 1993. Signal transduction mechanisms of recombinant bovine neurokinin-2 receptor stably expressed in baby hamster kidney cells. J. Cell Biochem. 52, 84–91.
- Ellis, J.L., Undem, B.J., Kays, J.S., Ghanekar, S.V., Barthlow, H.G., Buckner, C.K., 1993. Pharmacological examination of receptors mediating contractile responses to tachykinins in airways isolated from human, guinea pig and hamster. J. Pharmacol. Exp. Ther. 267, 95–101.
- Emonds-Alt, X., Vilain, P., Goulaoic, P., Van Broeck, D., Advenier, C., Naline, E., Neliat, G., Le Fur, G., Breliere, J.C., 1992. A potent and selective non-peptide antagonist of the neurokinin A (NK2) receptor. Life Sci. 50, 101–106.
- Erspamer, V., 1981. The tachykinin peptide family. Trends Neurosci. 4, 297–299.
- Evangelista, S., 2001. Involvement of tachykinins in intestinal inflammation. Curr. Pharm. Des. 7, 19–30.
- Fahy, J.V., Wong, H.H., Geppetti, P., Reis, J.M., Harris, S.C., Maclean, D.B., Nadel, J.A., Boushey, H.A., 1995. Effect of an NK1 receptor antagonist (CP-99,994) on hypertonic saline-induced bronchoconstriction and cough in male asthmatic subjects. Am. J. Respir. Crit. Care Med. 152, 879–884.
- Geppetti, P., Fusco, B., Marabini, S., Maggi, C.A., Fanciullacci, M., Sicuteri, F., 1988. Secretion pain and sneezing induced by the application of capsaicin to the nasal mucosa in man. Br. J. Pharmacol. 93, 509–514.
- Geppetti, P., Tramontana, M., Patacchini, R., Del Bianco, E., Santicioli, P., Maggi, C.A., 1990. Neurochemical evidence for the activation of the efferent function of capsaicin-sensitive nerves by lowering of the pH in the guinea-pig urinary bladder. Neurosci. Lett. 114, 101–106.
- Giuliani, S., Maggi, C.A., Rovero, P., Meli, A., 1988. Neurokinins induce a relaxation of the rat duodenum 'in vivo' by activating postganglionic sympathetic elements in prevertebral ganglia: involvement of a NK₂ type of neurokinin receptor. J. Pharmacol. Exp. Ther. 246, 322–327.
- Giuliani, S., Santicioli, P., Tramontana, M., Geppetti, P., Maggi, C.A., 1991. Peptide N-formyl-methionyl-leucyl-phenylalanine (FLMP) activates capsaicin-sensitive primary afferent nerves in guinea-pig atria and urinary bladder. Br. J. Pharmacol. 102, 730–734.

- Giuliani, S., Lecci, A., Giachetti, A., Maggi, C.A., 1993a. Tachykinins and reflexy-evoked atropine-resistant motility in the guinea-pig colon in vivo. J. Pharmacol. Exp. Ther. 265, 1224–1231.
- Giuliani, S., Santicioli, P., Lippe, I.T., Lecci, A., Maggi, C.A., 1993b. Effect of bradykinin and tachykinin receptor antagonist on xylene-induced cystitis in rats. J. Urol. 150, 1014–1017.
- Giuliani, S., Tramontana, M., Lecci, A., Maggi, C.A., 1996. Tachykinin receptors mediate atropine-resistant rat duodenal reflex contractions in vivo. Naunyn-Schmiedeberg's Arch. Pharmacol. 354, 327–335.
- Giuliani, S., Guelfi, M., Tolouse, M., Bueno, L., Lecci, A., Tramontana, M., Criscuoli, M., Maggi, C.A., 2001. Effect of a tachykinin NK₂ receptor antagonist, nepadutant, on cardiovascular and gastrointestinal function in rats and dogs. Eur. J. Pharmacol. 415, 61–71.
- Guard, S., Watson, S.P., 1991. Tachykinin receptor types: classification and membrane signalling mechanisms. Neurochem. Int. 18, 149–165.
- Hastrup, H., Schwartz, T.W., 1996. Septide and neurokinin A are high affinity ligands on the NK₁ receptor: evidence from homologus versus heterologus binding analysis. FEBS Lett. 399, 264–266.
- Henderson, A.K., Lai, J., Buck, S.H., Fujiwara, Y., Singh, G., Yamamura, M.S., Nakanishi, S., Roeske, W.R., Yamamura, H.I., 1990. A cloned NK₂ receptor mediates phosphatidylinositol hydrolysis in a transfected murine fibroblast. Life Sci. 47, PL7-PL12.
- Holzer, P., 1998. Tachykinins as targets of gastroenterological pharmacoterapy. Drugs News Perspect. 11, 394–401.
- Holzer, P., Holzer-Petsche, U., 1997a. Tachykinins in the gut: Part I. Expression, release, and motor function. Pharmacol. Ther. 73, 173–217.
- Holzer, P., Holzer-Petsche, U., 1997b. Tachykinins in the gut: Part II. Roles in neural excitation, secretion and inflammation. Pharmacol. Ther. 73, 219–263.
- Holzer, P., Lippe, I.T., Heinemann, A., Barthò, L., 1998. Tachykinin NK_1 and NK_2 receptor-mediated control of peristaltic propulsion in the guinea-pig small intestine, in vitro. Neuropharmacology 37, 131–138
- Hua, X.-Y., Theodorsson-Norheim, E., Lundberg, J.M., Kinn, A.-C., Hökfelt, T., Cuello, A.C., 1987. Co-localization of tachykinins and calcitonin gene-related peptide in capsaiicn-sensitive afferents in relation to motility effects on the human ureter in vitro. Neuroscience 23, 693–703.
- Joos, G., Van Schoor, J., Kips, J.C., Pauwels, R.A., 1996. The effect of inhaled FK 224, a tachykinin NK-1 and NK-2 receptor antagonist on neurokinin A-induced bronchoconstriction in asthmatics. Am. J. Respir. Crit. Care Med. 153, 1781–1784.
- Joos, G., Schelfhout, V., Van De Velde, V., Maggi, C.A., Pauwels, R.A., 2000. The effect of the tachykinin NK₂ receptor antagonist MEN 11420 (nepadutant) on neurokinin-A induced bronchoconstriction in patients with asthma. "Tachykinins 2000" La Grande Motte, France, October 17th–20th.
- Julia, V., Bueno, L., 1997. Tachykininergic mediation of viscerosensitive responses to acute inflammation in rats: role of CGRP. Am. J. Physiol. 272, G141–G146.
- Julia, V., Morteau, O., Bueno, L., 1994. Involvement of neurokinin 1 and 2 receptors in viscerosensitive response to rectal distension in rats. Gastroenterology 107, 94–102.
- Kiss, S., Lecci, A., De Groat, W.C., Maggi, C.A., Birder, L.A., 1999. The effect of the NK₂ receptor antagonist, MEN 11420, on proto-oncogene expression following experimental colitis. Soc. Neurosci. Abstr. 25, 411.
- Kraan, J., Vink-Klooster, H., Postma, D.S., 2001. The NK-2 receptor antagonist SR 48968C does not improve adenosine hyperresponsiveness and airway obstruction in allergic asthma. Clin. Exp. Allergy 31, 274–278.
- Kraneveld, A.D., James, D.E., De Vries, A., Nijkamp, F.P., 2000. Excitatory non-adrenergic, non-cholinergic neuropeptides: key players in asthma. Eur. J. Pharmacol. 405, 113–129.
- Krause, J.E., Chirgwin, J.M., Carter, M.S., Xu, Z.S., Hershey, A.D., 1987. Three rat preprotachykinin mRNAs encode the neuropeptides

- substance P and neurokinin A. Proc. Natl. Acad. Sci. U. S. A. 84, 881-885
- Krause, J.E., Staveteig, P.T., Mentzer, J.N., Schmidt, S.K., Tucker, J.B., Brodbeck, R.M., Bu, J.K., Karpitskiy, V.V., 1997. Functional expression of a novel human neurokinin-3 receptor homolog that binds (3H)senktide and (125-I-MePhe-7) neurokinin B, and is responsive to tachykinin peptide agonists. Proc. Natl. Acad. Sci. U. S. A. 94, 310–315.
- Ichinose, M., Nakajima, N., Takahashi, T., Yamauchi, H., Inoue, H., Takishima, T., 1992. Protection against bradykinin-induced bronchoconstriction in asthmatic patients by neurokinin receptor antagonists. Lancet 340, 1248–1251.
- Ichinose, M., Miura, M., Yamauchi, H., Kageyama, N., Tomaki, M., Oyake, T., Ohuchi, Y., Hida, W., Miki, H., Tamura, G., Shirato, K., 1996. A neurokinin 1-receptor antagonist improves exercise-induced airway narrowing in asthmatic patients. Am. J. Respir. Crit. Care Med. 153, 936–941.
- Ishizuka, O., Igawa, Y., Mattiasson, A., Andrsson, K.-E., 1994. Capsaicin-induced bladder hyperreactivity in normal conscious rats. J. Urol. 152, 525–530.
- Ishizuka, O., Mattiasson, A., Andrsson, K.-E., 1995. Prostaglandin E2-induced bladder hyperreactivity in normal conscious rats: involvement of tachykinins. J. Urol. 153, 2034–2038.
- Lai, J.P., Douglas, S.D., Ho, W.Z., 1998. Human lymphocyctes express SP and its receptor. J. Neuroimmunol. 86, 80–86.
- Laird, J.M., Olivar, T., Roza, C., De Felipe, C., Hunt, S.P., Cervero, F., 2000. Deficits in visceral pain and hyperalgesia of mice with a disruption of the tachykinin NK₁ receptor gene. Neuroscience 98, 345–352.
- Laird, J.M., Olivar, Lopez-Garcia, J.A., Maggi, C.A., Cervero, F., 2001.
 Responses of rat spinal neurons to distension of inflamed colon: role of tachykinin NK₂ receptors. Neuropharmacology, in press.
- Lecci, A., Maggi, C.A., 1995. Sensory neuropeptides in the lower urinary tract. In: Geppetti, P., Holzer, P. (Eds.), Neurogenic Inflammation. CRC Press, Boca Raton, Florida, USA, pp. 201–209.
- Lecci, A., Giuliani, S., Patacchini, R., Maggi, C.A., 1993. Evidence against a peripheral role of tachykinins in the initiation of micturition reflex in rats. J. Pharmacol. Exp. Ther. 264, 1327–1332.
- Lecci, A., Giuliani, S., Tramontana, M., Criscuoli, M., Maggi, C.A., 1997. MEN 11420, a peptide tachykinin NK₂ receptor antagonist, reduces motor responses induced by intravesical administration of capsaicin in vivo. Naunyn-Schmiedeberg's Arch. Pharmacol. 356, 182–188.
- Lecci, A., Giuliani, S., Tramontana, M., De Giorgio, R., Maggi, C.A., 1998a. The role of tachykinin NK1 and NK2 receptors in atropine-resistant colonic propulsion in anaesthetized guinea-pigs. Br. J. Pharmacol. 124, 27–34.
- Lecci, A., Giuliani, S., Tramontana, M., Santicioli, P., Criscuoli, M., Dion, S., Maggi, C.A., 1998b. Bladder distension and activation of the efferent function of sensory fibres: similarities with the effect of capsaicin. Br. J. Pharmacol. 124, 259–266.
- Lecci, A., Tramontana, M., Giuliani, S., Criscuoli, M., Maggi, C.A., 1998c. Effect of tachykinin NK₂ receptor blockade on detrusor hyperreflexia induced by bacterial toxin in rats. J. Urol. 160, 206–209.
- Lecci, A., De Giorgio, R., Barthò, L., Sternini, C., Tramontana, M., Corinaldesi, R., Giuliani, S., Maggi, C.A., 1999. Tachykinin NK₁ receptor-mediated inhibitory responses in the guinea-pig small intestine. Neuropeptides 33, 91–97.
- Lecci, A., Giuliani, S., Tramontana, M., Carini, F., Maggi, C.A., 2000. Peripheral actions of tachykinins. Neuropeptides 34, 303–313.
- Lördal, M., Navalesi, G., Maggi, C.A., Theodorsson, E., Hellstroem, P.M., 1999. The tachykinin NK-2 receptor antagonist nepadutant is a powerful inhibitor of small bowel motility stimulated by neurokinin A in man. Gastroenterology 116, A1032.
- Lundberg, J.M., Saria, A., 1982. Bronchial smooth muscle contraction induced by stimulation of capsaicin-sensitive sensory neurons. Acta Physiol. Scand. 116, 473–476.

- Lundberg, J.M., Brodin, E., Saria, A., 1983. Effects and distribution of vagal capsaicin-sensitive SP neurons with special reference to the trachea and lungs. Acta Physiol. Scand. 119, 243–252.
- Lundberg, J.M., Hokfelt, T., Martling, C.R., Saria, A., Cuello, C., 1984.
 Substance P immunoreactive sensory nerves in the lower respiratory tract of various mammals including man. Cell Tissue Res. 235, 251–261.
- Lunde, H., Hedner, J., Svedmyr, N., 1994. Lack of efficacy of 4 weeks treatment with the neurokinin receptor antagonist FK 224 in mild to moderate asthma. Eur. Respir. J. 7, 151S.
- Maggi, C.A., 1995. Tachykinins and calcitonin gene-related peptide (CGRP) as co-transmitters released from peripheral endings of sensory nerves. Prog. Neurobiol. 45, 1–98.
- Maggi, C.A., 1997. The effects of tachykinins on inflammatory and immune cells. Regul. Pept. 70, 75–90.
- Maggi, C.A., Schwartz, T.W., 1997. The dual nature of the tachykinin NK₁ receptor. Trends Pharmacol. Sci. 18, 351–355.
- Maggi, C.A., Abelli, L., Giuliani, S., Santicioli, P., Geppetti, P., Somma, V., Frilli, S., Meli, A., 1988a. The contribution of sensory nerves to xylene-induced cystitis in rats. Neuroscience 26, 709–723.
- Maggi, C.A., Santicioli, P., Patacchini, R., Cellerini, M., Turini, D., Barbanti, G., Beneforti, P., Rovero, P., Meli, A., 1988b. Contractile response of the human isolated urinary bladder to neurokinins: involvement of NK₂ receptors. Eur. J. Pharmacol. 145, 335–340.
- Maggi, C.A., Patacchini, R., Santicioli, P., Geppetti, P., Cecconi, R., Giuliani, S., Meli, A., 1989a. Multiple mechanisms in the motor responses of the guinea-pig isolated urinary bladder to bradykinin. Br. J. Pharmacol. 98, 619–629.
- Maggi, C.A., Santicioli, P., Geppetti, P., Parlani, M., Astolfi, M., Del Bianco, E., Patacchini, R., Giuliani, S., Meli, A., 1989b. The effect of calcium free medium and nifedipine on the release of substance P-like immunoreactivity and contractions induced by capsaicin in the isolated guinea-pig and rat bladder. Gen. Pharmacol. 20, 445–456.
- Maggi, C.A., Giuliani, S., Ballati, L., Lecci, A., Manzini, S., Patacchini, R., Renzetti, A.R., Rovero, P., Quartara, L., Giachetti, A., 1991a. In vivo evidence for tachykininergic transmission using a new NK-2 receptor-selective antagonist, MEN 10,376. J. Pharmacol. Exp. Ther. 257, 1172–1178.
- Maggi, C.A., Patacchini, R., Quartara, L., Rovero, P., Santicioli, P., 1991b. Tachykinin receptors in the guinea-pig isolated bronchi. Eur. J. Pharmacol. 197, 167–174.
- Maggi, C.A., Lecci, A., Santicioli, P., Del Bianco, E., Giuliani, S., 1992a. Cyclophosphamide cystitis in rats: involvement of capsaicin-sensitive primary afferents. J. Auton. Nerv. Syst. 38, 201–208.
- Maggi, C.A., Patacchini, R., Eglezos, A., Quartara, L., Giuliani, S., Giachetti, A., 1992b. Tachykinin receptors in the guinea-pig renal pelvis: activation by exogenous and endogenous tachykinins. Br. J. Pharmacol. 107, 27–33.
- Maggi, C.A., Patacchini, R., Rovero, P., Giachetti, A., 1993. Tachykinin receptors and tachykinin receptor subtypes. J. Auton. Pharmacol. 13, 23, 03
- Maggi, C.A., Catalioto, R.-M., Criscuoli, M., Cucchi, P., Giuliani, S., Lecci, A., Lippi, A., Meini, S., Patacchini, R., Renzetti, A.R., Santicioli, P., Tramontana, M., Zagorodnyuk, V., Giachetti, A., 1997. Tachykinin receptors and intestinal motility. Can. J. Physiol. Pharmacol. 75, 696–703.
- Maggio, J.E., 1988. Tachykinins. Annu. Rev. Neurosci. 11, 13-28.
- Martling, C.R., Saria, A., Fischer, J.A., Hokfelt, T., Lundberg, J.M., 1988. CGRP and the lung: neuronal coexistence with SP, release by capsaicin and vasodilatory effects. Regul. Pept. 20, 125–139.
- McLean, P.G., Picard, C., Garcia-Villar, R., Morè, J., Fioramonti, J., Bueno, L., 1997. Effects of nematode infection on sensitivity to intestinal distension: role of tachykinin NK₂ receptors. Eur. J. Pharmacol. 337, 279–282.
- Meini, S., Mak, J., Rohde, J.A.L., Rogers, D.F., 1993. Tachykinin control of ferret airways: mucus secretion, bronchoconstriction and receptor mapping. Neuropeptides 24, 81–89.

- Mussap, C.J., Geraghty, D.P., Burcher, E., 1993. Tachykinin receptors: a radioligand binding perspective. J. Neurochem. 6, 1987–2009.
- Nakanishi, S., 1987. Substance P precursor and kininogen: their structures, gene organizations and regulation. Physiol. Rev. 67, 1117–1142.
- Naline, E., Devillier, P., Drapeau, G., Toty, L., Bakdach, H., Regoli, D., Advenier, C., 1989. Characterization of neurokinin effects and receptor selectivity in human isolated bronchi. Am. Rev. Respir. Dis. 140, 679–686.
- Nawa, H., Hirose, T., Takashima, H., Inayama, S., Nakanishi, S., 1983.Nucleotide sequences of cloned cDNAs for two types of bovine substance P precursor. Nature 306, 32–36.
- Nawa, H., Kotani, H., Nakanishi, S., 1984. Tissue specific generation of two preprotachykinin mRNAs from one gene by alternative RNA splicing. Nature 312, 729–734.
- Palea, S., Corsi, M., Artibani, W., Ostardo, E., Pietra, C., 1996. Pharmacological characterization of tachykinin NK₂ receptors on isolated human urinary bladder prostatic urethra and prostate. J. Pharmacol. Exp. Ther. 277, 700–705.
- Parlani, M., Conte, B., Majmone, S., Maggi, C.A., Rovero, P., Giachetti, A., 1990. The contractile effect of tachykinins on human prostatic urethra: involvment of NK₂ receptors. J. Urol. 144, 1543–1545.
- Patacchini, R., Maggi, C.A., 1995. Tachykinin receptors and receptor subtypes. Arch. Int. Pharmacodyn. Ther. 329, 161–184.
- Patacchini, R., Santicioli, P., Zagorodnyuk, V., Lazzeri, M., Turini, D., Maggi, C.A., 1998. Excitatory motor and electrical effects produced by tachykinins in the human and guinea-pig isolated ureter and guinea-pig renal pelvis. Br. J. Pharmacol. 125, 987–996.
- Patacchini, R., Holzer, P., Maggi, C.A., 2000. Tachykinin autoreceptors in the gut. Trends Pharmacol. Sci. 21, 166.
- Patacchini, R., Cox, H.M., Stahl, S., Tough, I.R., Maggi, C.A., 2001. Tachykinin NK₂ receptor mediates contraction and ion transport in rat colon by different mechanisms. Eur. J. Pharmacol. 415, 277–283.
- Petersson, G., Malm, L., Ekman, R., Hakanson, R., 1989. Capsaicin evokes secretion of nasal fluid and depletes SP and CGRP from the nasal mucosa in the rat. Br. J. Pharmacol. 98, 930–936.
- Pietra, C., Bettellini, R., Hagan, R.M., Ward, P., McElroy, A., Trist, D., 1992. Effect of selective antagonists at tachykinin NK₁ and NK₂ receptors on xylene-induced cystitis in rats. Neuropeptides 22, 52.
- Regoli, D., Drapeau, G., Dion, S., D'orleans-Juste, P., 1989. Receptors for substance P and related neurokinins. Pharmacology 38, 1–15.
- Rupniak, N.M.J., Kramer, M.S., 1999. Discovery of the anti-depressant and anti-emetic efficacy of substance P receptor (NK₁) antagonists. Trends Pharmacol. Sci. 20, 485–489.
- Santicioli, P., Maggi, C.A., 1998. Myogenic and neurogenic factors in the control of pyeloureteral motility and ureteral peristalsis. Pharmacol. Rev. 50, 683-721.

- Sarau, H.M., Mooney, J.L., Schmidt, D.B., Foley, J.J., Buckley, P.T., Giardina, G.A.M., Wang, D.Y., Lee, J.A., Hay, D.W.P., 2000. Evidence that the proposed novel human "neurokinin-4" receptor is pharmacologically similar to the human neurokinin-3 receptor but is not of human origin. Mol. Pharmacol. 58, 552–559.
- Saria, A., Lundberg, J.M., Skofitsch, G., Lembeck, F., 1983. Vascular protein leakage in various tissues induced by SP, capsaicin, bradykinion, serotonin, histamine and by antigen challenge. Naunyn-Schmiedeberg's Arch. Pharmacol. 324, 212–218.
- Schuiling, M., Zuidhof, A.B., Zaagsma, J., Meurs, H., 1999. Roles of tachykinin NK₁ and NK₂ receptors in allergen-induced early and late asthmatic reactions, airway hyperresponsiveness, and airway inflammation in conscious unrestrained guinea-pigs. Clin. Exp. Allergy 29, 48–52.
- Stjärne, P., Lundblad, L., Lundberg, J.M., Angaard, A., 1989. Capsaicin and nicotine-sensitive afferent neurons and nasal secretion in healthy human volunteers and in patients with vasomotor rhinitis. Br. J. Pharmacol. 96, 693–701.
- Szolcsanyi, J., Barthò, L., 1982. Capsaicin-sensitive non-cholinergic excitatory innervation of the guinea-pig tracheobronchial smooth muscle. Neurosci. Lett. 34, 247–250.
- Takeda, Y., Krause, J.E., 1989a. Neuropeptide K potently stimulates salivary gland secretion and potentiates substance P-induced salivation. Proc. Natl. Acad. Sci. U. S. A. 86, 392–396.
- Takeda, Y., Krause, J.E., 1989b. γ-preprotachykinin-(72-92)-peptide amide potentiates substance P-induced salivation. Eur. J. Pharmacol. 161, 267–271.
- Tonini, M., Spelta, V., De Ponti, F., De Giorgio, R., D'Agostino, G.,
 Stanghellini, V., Corinaldesi, R., Sternini, C., Crema, F., 2001.
 Tachykinin-dependent and independent components of peristalsis in the guinea-pig isolated distal colon. Gastroenterology 120, 938–945.
- Toulouse, M., Coelho, A.M., Fioramonti, J., Lecci, A., Maggi, C.A., Bueno, L., 2000. Role of tachykinin NK₂ receptors in normal and altered rectal sensitivity in rats. Br. J. Pharmacol. 129, 193–199.
- Van Schoor, V., Joos, G.F., Chasson, B.L., Brouard, R.J., Pauwels, R.A., 1998. The effect of the NK2 tachykinin receptor antagonist SR 48968 (saredutant) on neurokinin A induced bronchoconstriction in asthmatics. Eur. Respir. J. 12, 17–23.
- Zagorodnyuk, V., Maggi, C.A., 1995. Neuronal tachykinin NK₂ receptors mediate release of nonadrenergic noncholinergic inhibitory transmitters in the circular muscle of guinea-pig colon. Neuroscience 69, 643–650.
- Zhang, Y., Lu, L., Furlonger, C., Wu, G.E., Paige, C.J., 2000. Hemokinin is a hematopoietic-specific tachykinin that regulates B lymphopoiesis. Nat. Immunol. 1, 392–397.